

Management of *Ralstonia* wilt of tomato through microbes, plant extract and combination of cake and chemicals

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ABSTRACT: The experiment was conducted on wilt of tomato (*Lycopersicon esculentum* L.), caused by *Ralstonia solanacearum* Yabuuchi, during post monsoon season for four consecutive years (1998-99 to 2002-03) to record the impact of three groups viz; (i) microbes (ii) plant extract (iii) cake and chemicals on the yield of tomato as well as on the viability of inocula. The pooled data revealed that there was significant effect in all the treatments in reducing the primary inocula. Microbes viz; *Glomus mosseae*, *Trichoderma viride* and *Azotobacter+Phosphobactrin* resulted in reduction of primary inocula (41.4, 29.0 & 37.3% at 90 days over initial population, respectively) of *Ralstonia solanacearum* in soil and increased the yield by 141.1, 142.1 & 89.9%, respectively. Similar trend was observed in plant extracts viz; Asafoetida +Turmeric, Onion and Garlic in which the yield increase was noted to be 71.2, 71.2 & 64.4%, respectively over control. Karanj cake along with bleaching powder was more effective in reducing the apparent bacterial growth rate at flowering stage and increased the yield by 123.4% followed by bleaching powder and lime 107.5% over control.

Key words: *Ralstonia* wilt, tomato, microbes, cake and chemicals, plant extracts

Soil is a heterogeneous habitat of beneficial, plant pathogenic and saprophytic microbes, which plays an important role in the spread of soil borne diseases. *Ralstonia solanacearum* Yabuuchi is the causal agent of wilt of solanaceous vegetables (Samaddar *et al.*, 1998). Among them, tomato (*Lycopersicon esculentum*) is one of the important vegetables, which suffers badly from this disease in summer, rainy and winter seasons. Infested soil and surface water, irrigation water, are the main sources of spread of inocula. The pathogen can infect undisturbed roots of susceptible hosts through microscopic wounds caused by the emergence of lateral roots. After infection, the bacterium colonizes the cortex and makes its way towards the xylem vessel; from there it rapidly spreads throughout the plant (Momol *et al.*, 2005). It has been reported that infected roots present in soil released vast number of *R. solanacearum* cells into the rhizosphere and invades subsequent crops to cause of spread the disease (Mc Carter, 1976). The race 1 of *R. solanacearum* can survive for more than 6 months

on seed surface and in the soil (Samaddar *et al.*, 1998) and also reported to be spread through seed transmission (Singh *et al.*, 1995). The meager information on management by microbes (Kobayaashi, 1999), plant extracts (Bora *et al.*; 1997), cakes (Sharma and Kumar, 2000) and chemicals (Michel *et al.*, 1997; Mazumder, 1998) are available. Hence the study was undertaken on effect of microbes, plant extracts, cake and chemicals on primary inoculums and impact of yield reported here under.

MATERIALS AND METHODS

The experiment was conducted in R.B.D. with 3 replications during post monsoon seasons of 1998-99 to 2002-03 on cv. Pusa Ruby with treatments; (i) microbes (a) mycorrhiza (*Glomus mosseae*) @500 g/m² of dry root mass multiplied in finger millet (ragi) *Penisetum typhoides* in pot (b) *Trichoderma viride* @5 kg/ha mixed in FYM (c) *Azotobacter chroococcum* + *Phosphobactrin* @2 kg each mixed in 50 kg FYM + 50 kg soil (ii) Plant extracts viz; (a) asafoetida (*Ferula foetida*) @1 g +

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turmeric (*Curcuma longa*) @5 g in 10 litres water (b) onion (*Allium cepa*) @5% extract (c) garlic (*Allium sativum*) @ 5% extract (iii) cake and chemicals viz; (a) karanj (*Deris indica*) cake @10 q/ha + bleaching powder (CaOCl_2) @30 kg/ha and (b) bleaching powder (CaOCl_2) @30 kg/ha + lime @2500 kg/ha (c) urea @200 kg/ha + lime (CaCO_3) @5000 kg/ha (v) control. The microbes as well as cake and chemicals (viz; bleaching powder and lime) were applied 15 days earlier to transplanting. The plant extracts were used for seedling dip for 30 minutes and sprayed three times at 10 days interval after one month of transplanting. Twenty-one days seedlings were transplanted and recommended doses of NPK were applied. The soil samples were collected at 0, 30, 60 & 90 days and the bacterial count was made on TTC media (Kelman, 1954) by dilution method and data were statistically analysed. The apparent bacterial growth rate (r) was calculated by following the formula given below after conversion of bacterial counts at different days in to LOGIT value described by Van der Plank (1963) and reported in here with:

$$r = 1/t_2 - t_1 [\log_e (X_2 / X_1)]$$

Where, X_1 = initial count (cfu x 10^4 /g soil)

X_2 = final count (cfu x 10^4 /g soil)

The apparent infection rate was presented graphically. The yield recorded year wise were pooled, statistically analyzed and presented herewith.

RESULTS AND DISCUSSION

Effect of microbes

Mycorrhiza (*Glomus mosseae*): It is revealed from the mean of four years (1998-99 to 2002-03) data that there was a significant effect of treatments on reduction of *Ralstonia solanacearum* population in soil. Addition of VAM mycorrhiza (*Glomus mosseae*) results in maximum reduction (41.4%) of *Ralstonia solanacearum* population. The VAM was effective at 30 days in which the apparent bacterial growth rate (BGR) was ($r = -0.032$ unit/day), at flowering stage ($r = -0.011$ unit/day) at 60 days interestingly later on it reduced at 90 days ($r = -0.009$ unit/day) (Fig.1). The trend resulted in a

Table 1. Effect of microbes, plant extracts and cake and chemicals on primary inoculum of bacterial wilt of tomato cv. Pusa Ruby (1998- 99 to 2002-03)

Treatments	<i>Ralstonia solanacearum</i> (cfu x 10^4 /g soil) at days					Av of 30,60 & 90 day	% decrease by 90 days	% decrease over control	Yield* (g/ha)	% Increase over control
	Initial	30	60	90	Av of					
Microbes	<i>Glomus mosseae</i>	84.4	64.4	56.4	49.5	56.8	41.4	39.41	76.18	141.1
	<i>Trichoderma viride</i>	83.3	61.0	70.2	59.2	63.5	29.0	27.52	76.50	142.1
	<i>Azotobacter</i> + <i>Phosphobactrin</i>	81.5	72.2	56.1	59.1	59.8	37.3	27.62	60.00	89.9
Plant extracts	Asafoetida + turmeric	75.3	69.0	53.0	45.7	55.9	39.3	44.01	54.10	71.2
	Onion extract	70.8	77.5	52.3	50.5	60.1	32.7	38.09	54.10	71.2
	Garlic extract	97.4	76.1	72.9	73.6	74.2	24.4	9.80	51.95	64.4
Cake & chemicals	Karanj cake + bleaching powder	78.8	67.0	56.3	53.2	58.8	32.5	34.87	70.60	123.4
	Bleaching powder + lime	80.1	61.1	64.8	51.1	59.0	36.2	37.37	65.58	107.5
	Urea +lime	91.2	63.8	65.6	63.1	63.5	30.8	22.72	48.23	52.6
Control		81.6	72.2	55.4	54.2	60.6	33.6	-	31.60	
CD(P 0.05)					6.1			260.17		
CV (%)								30.66		

*Mean of four years (1998-99 to 2001-02)

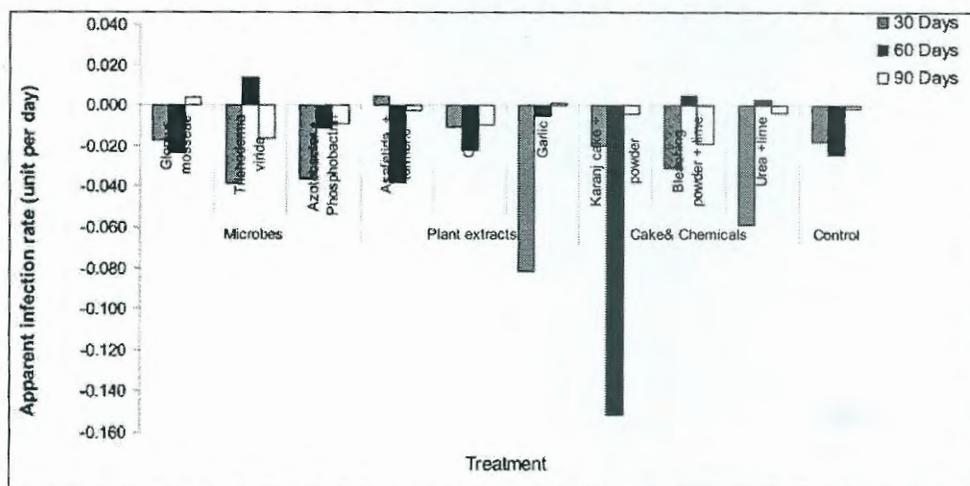


Fig. 1. Effect of microbes, plant extract cake and chemical on apparent infection rate of *Ralstonia solanacearum* in tomato cv Pusa Ruby

logarithmic reduction in apparent infection rate (Fig.2). This effect of reduction may be due to colonization by the *G. mosseae* in the root. It has been reported that Vascular Arbuscular Mycorrhiza (VAM) and charcoal compost caused reduction in the bacterial wilt in tomato (Kobayashi, 1990). The extract of tomato roots colonized with *G. fasciculatum* reduced population of *P. solanacearum* in nutrient broth *in vitro* (Suresh and Rai, 1991). Several reports on VAM fungi are available that these fungi decrease the severity of many soil borne fungal diseases as well as that caused by *P. solanacearum*, however; further studies are required to establish the mechanism of active principle of VAM fungi inhibiting *P. solanacearum*. (Bagyaraj, 2002).

The yield was found maximum 760.18 kg/ha which was 141.1% increase over control (310.6 kg/ha). The increase in yield may be due to mycorrhizal symbiosis which might have colonized with roots resulting into higher plant uptake of water and minerals from the soil thereby the yield has increased. It has been reported that mycorrhizal fungus colonizes the cortical cell of the roots, thereby form a symbiotic relationship with the host (Read *et al.*, 1992).

Trichoderma viride

Trichoderma viride supplemented soil resulted in 29.0% reduction at 90 days the in *R. solanacearum* population. The apparent infection

rate was ($r = -0.039$ unit /day) at 30 days and ($r = -0.016$ unit/day) at 90 days (Fig.1). The trend resulted in logarithmic reduction in apparent infection rate (Fig.2). It has been reported that *Trichoderma* spp. have ability to adopt extreme soil conditions viz, soil temperature, moisture and fungicidal load added in soil and that is why, it is importance in the field of biological control of plant diseases. The *T. viride* produced antibiotics viz, viridin, dermadine and gliotoxin (Sawant, 2002). A vast number of plant pathogens, particularly soil borne fungi controlled by *Trichoderma* spp. (Anahosur, 1999). Singh *et al.* (2004) has reviewed and reported that *Trichoderma*, a microbe has multifaceted activity.

Application of *T. viride* in soil resulted 760.5 kg/ha which was 142.1% increase over control (Fig.2). *T. viride* used as bioagent for control of various fungal pathogens viz; *Macrophomina* stem rot and root rot in Sesame (Rajpurohit, 2004), white rot in Pea (Kapoor and Kumar, 2004), Seed borne pathogen in Okra (Gurjar *et al.*, 2004) and bacterial wilt in chilli (Yan Sihuang *et al.*, 2005).

Azotobacter chroococcum

Azotobacter chroococcum and *Phosphobactrin* resulted in reduction (37.3%) at 90 days in the primary inoculum. The apparent bacterial growth rate was $r = -0.018$ unit/ day at 30 days and at flowering stage (60 days) was $r = -0.011$ unit /day (Fig.1) and remained ineffective in reducing ($r =$

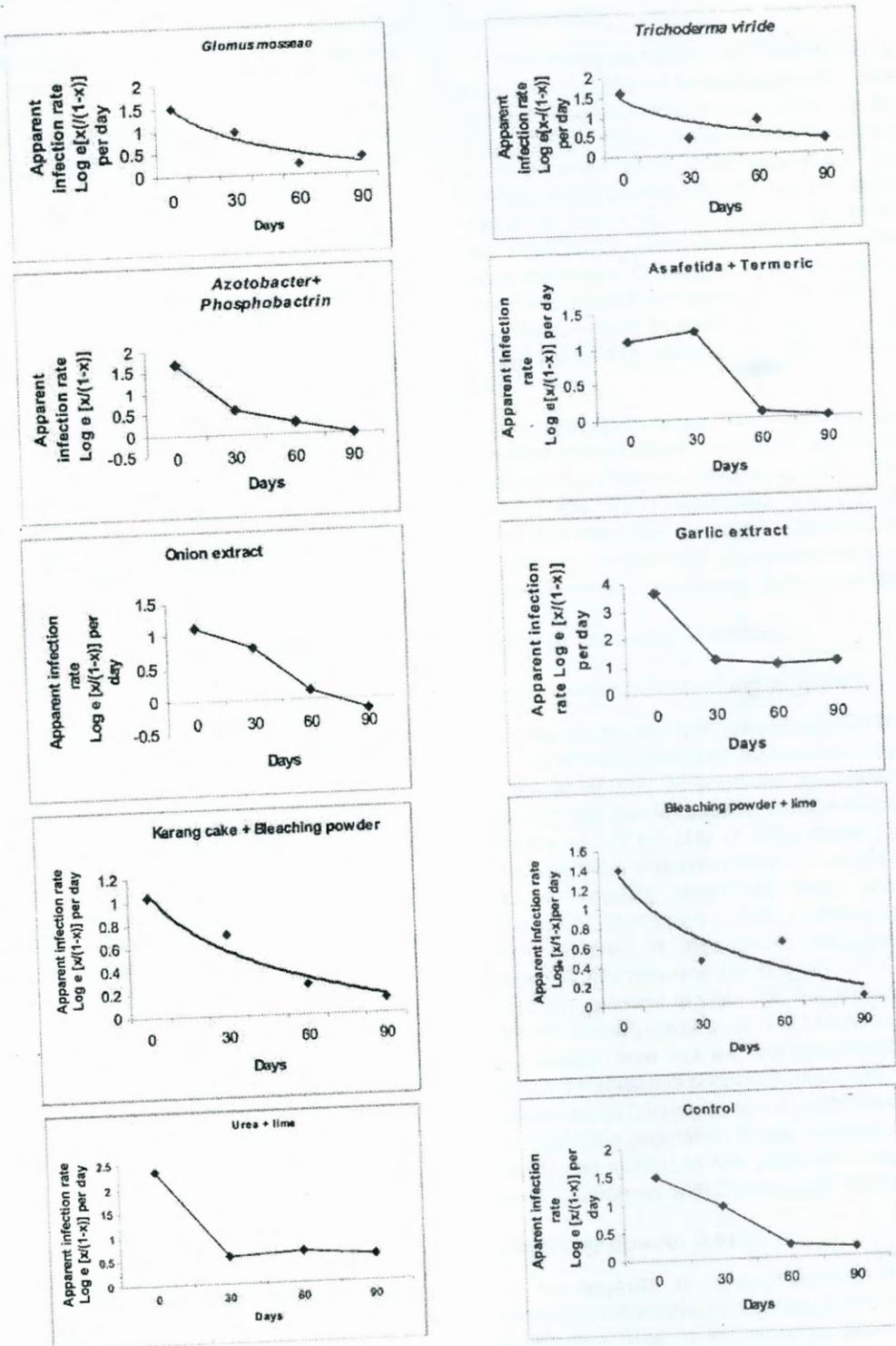


Fig. 2. Apparent infection rate (unit /day) under various treatments

0.004 unit/day) at 90 days. The trend showed a logarithmic reduction in apparent infection rate (Fig. 2). It has been reported that extract of tomato roots colonized with N-fixing bacteria *Azotobacter* was ineffective (Suresh and Rai, 1991). It is reported that the *Azotobacter* has ability to produce anti-fungal antibiotics and also to fix nitrogen, which improve the fertility of soil (Laxmi – Kumari *et al.*, 1975) and also act as bioagents (Wani, 2002). Thus the results indicated that the use of microbes will be effective for longer period in managing the *R.solanacearum* population in soil. *Azotobacter chroococcum* treated plot resulted in yield of 600.0kg/ha which was 89.9% increase over control.

Effect of plant extract: asafoetida and turmeric

The asafoetida + turmeric powder treated plot also reduced the primary inocula to 32.7% at 90 days. In the early stage (30 days) the effectiveness was slow. The apparent infection rate was $r = -0.038$ unit/day at flowering stage (60 days) and it remain effective in reducing at 90 days ($r = -0.002$ unit /day). The trend indicated moving trend which was different from microbes (Fig.2). It has been reported that seedling treatment with mixture of asafoetida (*Ferula foetida* Ritz.), turmeric powder (*Curcuma longa* Linn.) and water (ATW) in ratio of 1g: 5g: 10 liters controlled bacterial wilt or by spraying 5% ATW solution or soil drenching at 15, 30 and 45 days after transplanting (Bora, 1995, Pun and Das, 1997). In the present investigation asafetida + turmeric powder treated plot resulted 540.1 kg/ha which was 71.2% increase over control. The result was in conformity with Mazumder (1998) reported from Assam.

Onion extract

The seedling treated with onion extracts (5%) a significant reduction in primary inoculum was also found to be 39.3% at 90 days. The apparent infection rate was $r = -0.0106$ unit/day at 30 days whereas $r = -0.023$ unit/day at flowering stage (60 days) and $r = -0.01$ unit/day at 90 days (Fig.1). The trend was also different from microbes at 30 days and reduction was more at 60 days Fig 2. Thus the plant extracts can also be used for minimizing the *Ralstonia solanacearum* population in soil, which will be cheaper than other treatment. The onion extract treated plot resulted in yield of 540.1 kg/ha which was 71.2% increase over control. Onion extract was better than garlic extract.

Garlic extract

The seedlings treated with garlic extracts (5%) were less effective in reducing primary inoculums which was also found to be 9.97% at 90 days (Fig.1). The seedling treated with garlic extract (5%) showed more effective at the initial stage (30 days) in which the apparent infection rate $r = -0.082$ unit/day at 30 days and $r = -0.01$ unit/day at 60 days and later on its effectiveness was reduced. The trend was also different from microbes but the reduction rate was constant after 30 days. (Fig.2). The garlic extract treated plot resulted in yield of 510.95 kg/ha which was 64.4% increase over control only.

Several workers have reported that the extract of onion (*Allium cepa*) and garlic (*Allium sativum*) reduced the disease (Hutaglun, 1988; Hanudin, 1987; Komarova and Korunets, 1997) *in vitro* and *vivo*. Thus the plant extracts can also be used for minimizing the *Ralstonia solanacearum* population in soil, which will be cheaper than other treatment.

Effect of cake and chemical

Karanj cake and bleaching powder

The soil treated with cake along with bleaching powder (CaOCl_2) reduced the primary inoculum to 32.5% at 90 days whereas the apparent infection rate was effective in reducing the apparent infection rate $r = -0.021$ unit/day at initial stage (30 days). The maximum reduction rate $r = -0.152$ unit/day was at flowering stage (60 days) and remain effective at 90 days $r = -0.004$ unit/day (Fig.1). The trend resulted in logarithmic reduction in the apparent infection rate per day (Fig.2). Cake and Chemical resulted in yield 700.6 kg/ha which was 123.4% increase over control. It has been reported that karanj cake act as organic amendment to reduce the *Ralstonia* population in soil and bleaching powder act as bactericide which might have reduced the *Ralstonia* population in soil resulting into good health and increased the yield. The above result was in conformity with Sharma and Kumar (2000).

Bleaching powder and lime

As regards to chemicals, the maximum reduction in bacterial population in soil was 36.2% at 90 days (Fig. 1) in bleaching powder + lime treated plot. The apparent BGR $r = -0.031$ unit/day was at initial stage (30 days) and at final stage r

= -0.019 unit/day at 90 days. The trend resulted in logistic reduction in the apparent infection rate per day (Fig.2). The yield obtained was 650.58 kg/ha which was 107.5% increase over control. It has been reported that application of CaO alone significantly reduced the population of *Ralstonia* (Michel *et al.*, 1997). Pre plant application of lime 200 lb/acre in form of CaCO₃ augmented Ca accumulation in leaf tissue and soil and reduced the rate of bacterial development on susceptible cultivars soil but had little effect on fruit yield (Locscio *et al.*, 1998). Yamazaki *et al.* (1998) have reported that the resistance was negated at low concentration in Hawaii 7998 (highly resistant) and pathogen population in stem decreased with increasing Ca concentration. On the contrary to above Jaworski and Morton (1964) did not found any difference in plant survival in a manorial experiment with 4 tomato varieties, and the plants receiving different levels of Ca and Mg.

Urea and lime

Urea (200 Kg/ha N) and CaCO₃ (5000 Kg/ha) treated plot resulted reduction in primary inoculums 30.8% at 90 day which was significant reduction in BGR. The apparent BGR(r) = -0.059 unit/day at 30 days and r = -0.004 unit/day at 90 days. The apparent infection rate was different from other two combined above treatments (Fig.2). The yield was increased to 52% over control.

The relative impact of microbes, plant extracts, cake and chemical with respect to *Ralstonia solanacearum* population and yield were presented above clearly indicated that the yield increased maximum to 142.1% and 141.1%, respectively with use of *Trichoderma viride* and Mycorrhiza i.e. *Glomus mosseae* than control. However, karanj cake along with bleaching powder was more effective in reducing the apparent bacterial growth rate at flowering stage, which is the most critical stage of bacterial wilt, in which 123.2% more yield was obtained which was at par with that of microbes than control whereas the plant extract resulted more effective in reducing the *Ralstonia solanacearum* population in soil. Thus microbes, Karanj cake and bleaching powder resulted in logistic reduction in primary inocula which are sustainable in management of primary inocula of *Ralstonia solanacearum* in soil and increased the yield.

REFERENCES

- Anahosur, K.H. (1999). Management of plant disease through antagonist. An over view. In: *Recent advances in plant pathology* (Eds. Vaidya, J. G. and S.Y. Kamble.) Dept. of Botany, Univ. of Pune pp. 38-56.
- Bagyaraj, D.J. (2002). VA Mycorrhiza: Utility in the control of plant pathogens. In ICAR Sponsored Winter School on "Frontiers in ecofriendly management of crop diseases and pests and utility of biofertilizers in improving soil fertility" (Aug.6 to Sept. 4). Department of Plant Pathology and Agriculture Microbiology. Mahatma Phule Krishi Vidyapeeth, Rahuri. pp. 37-41.
- Bora, L.C., Gogoi, P.K. and Samuel, Jaya (1997). Use of botanicals in management of bacterial wilt of tomato caused by *Pseudomonas solanacearum* (Abstr.) In *Symposium on Major Diseases of Crop Plants in Eastern India and Their Management*. Dec.10-11. Assam Agril Univ. pp. 11(Mimeograph).
- Bora, L.C. (1995). Management of bacterial wilt of tomato by organic formulation. *Plant Health* 1: 74-6.
- Gurjar, K.L. Singh, S.D. and Rawal, P. (2004). Management of seed borne pathogen of okra with bioagents. *Plant Disease Research* (Ludhiana) 19(1) 44-46. *Fide Rev. Plant Path.* (2005) 84: 350. pp. 57.
- Hanudin (1987). Controlling the incidence of bacterial wilt (*Pseudomonas solanacearum* E.F.Smith) on tomato plants by some plant extracts. *Bulletin Peneltian Horticulture*. 15: 60-6. *Fide Rev. Plant Path.* (1991) 70: 2249.
- Hutagalung, L. (1988). Garlic bulb as a material for suppressing the incidence of bacterial wilt (*Pseudomonas solanacearum* E.F.Smith) on tomatoes. *Bulletin Peneltian Hort. Kultura* 16: 84-93 *Fide Hort. Abstract* (1990) 60: 3477. *Rev. Plant Path.* (1991) 70: 6778.
- Jaworski, C.A. and Morton D.J. (1964). An epiphytic of *Pseudomonas solanacearum* in tomatoes on newly-cleared Klej sand in relation to potassium, calcium and magnesium levels. *Pl. Dis. Rep.* 48: 88-9. *Fide Hort. Abstract* (1964) 34: 5047.
- Locascio, S.J., Stall, R.E. and Stall, W.M. (1998). Bacterial wilt expression in tomato as influenced by cultivars and lime. *Proceeding of the Florida State Horticultural Society* 101: 356-8. *Rev. Plant Path.* (1952) 71: 4204.
- Kapoor A.S. and Pradeep Kumar (2004). Evaluation of different organic substrate, viability of

- Trichoderma harzianum* and management of white rot of pea. *Plant Disease Research* (Ludhiana) 19(1) 55-56. *Rev. Plant Path.* (2005) 84: 133. pp. 22.
- Kelman, A. (1954). The relationship of pathogenicity in *Pseudomonas solanacearum* to colony appearance on tetrazolium medium. *Phytopathology* 44: 693-5.
- Kobayashi, N. (1990). Biological control of soil borne diseases with VAM fungi and charcoal compost. In: *The Biological Control of Plant Disease*. Proceeding on the International Seminar "Bio Control of Plant Diseases and Virus vector's held in Tsukuba, Japan, Sept. 17-21. (Edited by Komada, H.; Kiritani, K.; Bay- Peterson, J.), *Rev. Plant Path.* (1992) 71: 7429.
- Komarova, M.S. and Korunets, I.V. (1997). Biological means for control of bacterial disease of tomatoes. *Zashchita Karantin Rastenii* No. 4, 27. *Rev. Plant Path.* (1998) 77: 2239.
- Laxmi-Kumari, Vijaya Laxmi, M.K. and Subba, Rao, N.S.S. (1975). *Phytopath.* 75: 27-30.
- Mazumder, N. (1998). Managing *Ralstonia solanacearum* wilt in tomato. *Journal of Mycology and Plant Pathology* 28: 189-92. *Rev. Plant Path.* (1999) 78: 1361.
- Mc Carter, S.M. (1976) Persistence of *Pseudomonas solanacearum* in artificially infected soils. *Phytopathology* 66: 998-1000. *Hort. Abstract* (1977) 47: 3702.
- Michel, V.V., Wang, J.F., Midmore, D.J. and Hartman, G.L. (1997). Effect of intercropping and soil amendments with urea and calcium oxide on the incidence of bacterial wilt of tomato and survival of soil borne *Pseudomonas solanacearum* in Taiwan. *Plant Disease* 46: 600-10. *Rev. Plant Path.* (1997) 76: 9882.
- Momol, Tim, Pingsheng Ji, Ken Pernezny, Robert McGovern and Steve Olson (2005). Soil borne tomato diseases caused by *Ralstonia* and *Fusarium* species and their field diagnostics (A document is Fact Sheet PP-205, one of a series of the Plant Pathology Department, Florida Cooperative Extension Service, Institute of Food and Agricultural Sciences, University of Florida).
- Pun, K.B. and Das, G.R. (1997). Management of bacterial wilt asafetida and turmeric. *Bacterial wilt Newsletter*. No.14, 6. *Rev. Plant Path.* (1998) 77: 515.
- Rajpurohit, T.S. (2004). *Trichoderma viride*: Bio-control agent effective against *Macrophomina* stem rot and root rot of sesame. *Journal of Eco-Physiology* 7(3/4) 187-188. *Review of Plant Path.* (2006) 85: 4519. pp. 725.
- Read, D.J., Lewis, D.H., Fitter, A.H. and Alexander, I.J. (1992). Mycorrhizas in ecosystem. *CAB International*, Walling Ford, London.
- Samaddar, K.R., Chakraborty, M. and Kanjilal, S. (1998). Identification of the race 1 of *Pseudomonas solanacearum* causing solanaceous vegetables in West Bengal and its survival. *J. Mycopathol. Res.* 36: 51-58.
- Sawant, D.M. (2002). *Trichoderma*: A potent plant disease control agent. In ICAR Sponsored Winter School on "Frontiers in ecofriendly management of crop diseases and pests and utility of biofertilizers in improving soil fertility" (Aug.6 to Sept. 4). Department of Plant Pathology and Agriculture Microbiology. *Mahatma Phule Krishi Vidyapeeth*, Rahuri. pp. 14-16.
- Sharma, J.P. and Kumar, S. (2000). Management of *Ralstonia* wilt through soil disinfectant, mulch, lime and cakes in tomato (*Lycopersicon esculentum*). *Indian J. Agril. Science* 70: 17-19.
- Singh, U.S., Zaidi, N.W., Joshi, D., Khan, T., John, D. and Bajpai, A. (2004). *Trichoderma*: A microbes with multifaceted activity. In *Annu. Rev. Pl. Pathol.* Vol. 3. pp. 33-75.
- Singh, B., Anita, K., Sharma, K.D., Agrawal, P.C. and Ramnath (1995). *World distribution of phytopathogenic bacteria*, NBPGR, New Delhi. pp. 147.
- Suresh, C.K. and Rai, P.V. (1991). Interaction of *Pseudomonas solanacearum* with antagonistic bacterium and VAM mycorrhiza. *Current Research University of Agriculture Science* 20(3): 36-7. *Rev. Plant Path.* (1993) 72: 1016.
- Wani, P.V. (2002). Biofertilizers as bio-agents. In ICAR Sponsored Winter School on "Frontiers in ecofriendly management of crop diseases and pests and utility of biofertilizers in improving soil fertility." (Aug.6 to Sept. 4). Department of Plant Pathology and Agriculture Microbiology. *Mahatma Phule Krishi Vidyapeeth*, Rahuri. pp. 27-36.
- Van Der Plank, J.E. (1963). *Plant Diseases: Epidemics and Control*. Academic Press, New York, pp. 349.
- YanSihuang, Wu Siping, Lu Deqing and Chen Xiaojun (2005). Effects of the biological control strain of *Trichoderma harzianum* on microflora in rhizosphere. *Southwest China Journal of Agriculture Sciences* 18(1): 40-46. *Rev. Plant Path.* (2006) 85: 2866. pp. 460.
- Yamazaki, H., Kikuchi, S., Hoshima, T. and Kimura, T. (1995). Effects of compost application on bacterial wilt (*Ralstonia solanacearum*) development and calcium uptake of tomato seedling. *Japanese Journal of Soil Science and Plant Nutrition* 69(1): 78-9. *Rev. Plant Path.* (1998) 77: pp. 9325.